Biosynthesis of Ether-Type Polar Lipids in Archaea and Evolutionary Considerations

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INTRODUCTION

The concept of Archaea was proposed on the basis of phylogenetic analysis of small-subunit rRNA sequences (109). In addition to the rRNA sequences, a number of biochemical properties of Archaea that are distinct from those of Bacteria and Eucarya supported the concept that these three domains are the most basic taxon of all living organisms. Membrane polar lipids are some of the most remarkable features among the distinct characteristics of archaea. Archaeal lipids have been studied since the 1960s from the structural and biosynthetic points of view. Throughout this period, a great number of novel and unique structures of archaeal polar lipids were reported and reviewed (19, 47, 49, 52, 53). Regarding these structures, we recognized four structural characteristics of archaeal polar lipids: (i) The stereostructure of the glycerophosphate backbone: hydrocarbon chains are bound at the sn-2 and sn-3 positions of the glycerol moiety in archaeal lipids, while bacterial and eucaryal lipids have sn-1 and 2-diradyl chains. That is, the glycerophosphate backbone of archaeal phospholipids is sn-glycerol-1-phosphate (G-1-P), which is an enantiomer of the sn-glycerol-3-phosphate (G-3-P) backbone of bacterial and eucaryal phospholipids. (ii) Ether linkages: hydrocarbon chains are bound to the glycerol moiety exclusively by ether linkages in...
archaeal lipids, in contrast to the situation for bacterial polar lipids, most of which have ester linkages between fatty acids and a glycerol moiety. (iii) Isoprenoid hydrocarbon chains: hydrocarbon chains of polar lipids are highly methyl-branched isoprenoids in archaea, while their bacterial counterparts are mostly straight-chain fatty acids. (iv) Bipolar tetraether lipids: bipolar lipids with a tetraether core are present in significant numbers in archaeal species. These tetraether polar lipids span the membrane to form a membrane monolayer. Among these characteristics, the stereostructure of the glycerophosphate backbone is the most specific to organisms of each domain in terms of structure. It is the most crucial feature of archaeal lipids of phylogenetic and evolutionary significance (see below). However, the enantiomeric difference appears to be insignificant for the physicochemical properties of the lipid membrane of archaea, because the enantiomers are equivalent in physicochemical properties except for chirality. The ether linkages, isoprenoid chains, and bipolar tetraether lipids are significant for the physicochemical properties of archaeal lipid membranes, for example, their phase behavior and the permeability of the membranes.

In contrast to the four differences, most of the polar head groups of phospholipids are shared by organisms of the three domains, with minor exceptions. Ethanolamine, l-serine, glycerol, myo-inositol, and even choline are found throughout the three domains as phosphodiester-linked polar head groups in phospholipid (31, 52, 53). Some minor unique polar groups, such as di- and trimethylaminopentanetetrols (30), glucosaminyl-myoinositol (78), and glucosyl-myoinositol (71), are found in a limited number of species of archaea.

The biosynthetic mechanisms by which these characteristic lipid structures are formed have been a subject of interest for a long time. As early as 1970, the first metabolic study on archaeal lipids was reported (50). It comprised in vivo incorporation experiments of radioactive glycerol into lipids in *Halobacterium cutirubrum* (later renamed *Halobacterium salinarum* [104]), which allowed exploration of the mechanism of the formation of the enantiomeric glycerophosphate backbone. However, only in vivo studies were reported occasionally in the 20 years after 1970. In 1990, the first in vitro study of archaeal lipid biosynthesis was published (115). The enzymatic activity of ether bond formation was reported for *Methanobacterium thermoautotrophicum* strain Marburg (recently reclassified *Methanothermobacter marburgensis* [107]). In vitro studies of the major pathway of polar lipid biosynthesis in archaea have been published in the 15 years since the in vivo studies were carried out during the preceding two decades (1970 to 1990). To the best of our knowledge, no review specifically dealing with the in vitro biosynthesis of archaeal lipids has yet been published. Recently, phylogenetic analyses and evolutionary considerations of the enzymes for lipid biosynthesis were carried out (10), but polar lipid biosynthesis was not discussed. Although previous reviews mainly dealing with archaeal lipid structures 10 years or more ago partly described biosynthetic aspects, those were restricted to in vivo studies (48, 53). Therefore, this is the first review that focuses mainly on in vitro biosynthetic studies of archaeal polar lipids. The present review assembles collected knowledge on the recent progress in in vitro studies of the biosynthesis of the four characteristic structures of polar lipids in archaea along with the preceding in vivo studies (isoprenoid biosynthesis, sn-glycerol-1-phosphate formation, ether bond formation, phospholipid polar head group attachment, glycolipid synthesis, and tetraether lipid formation), and it also provides a comparison with bacterial polar lipid biosynthesis.

In spite of the limited number of enzymatic studies of polar lipid biosynthesis in archaea, quite a number of genes relevant to lipid biosynthesis have been found in the genome sequences...
of archaea. Homology searches for enzymes that are expected to be involved in archaeal polar lipid biosynthesis have revealed many homologs, which have been used for the detection of activities of unknown enzymes and for the evolutionary analysis of the polar lipid synthetic pathway. The evolutionary aspects of polar lipid biosynthesis are also discussed in this review. Because polar lipids are the principal and essential constituents of the cytoplasmic membranes of all cells, fundamental differences in polar lipid structure likely reflect the history of diversification of fundamental cellular lines. The uniformity and diversity of the membrane polar lipid structures of archaea and bacteria are discussed. The nomenclature for archaeal polar lipids proposed by Nishihara et al. (77) is used throughout this paper. Scientific names of archaea as they appear in the original references or have been changed most recently are used in this text. The changes in nomenclature of the archaea appearing in the present text are listed in Table 1.

## ISOPRENOID BIOSYNTHESIS

The hydrocarbon portions of archaeal diether lipids are exclusively isoprenoids (C<sub>30</sub> phytanyl, C<sub>25</sub> sesterterpenyl or farnesylgeranyl groups). While isoprenoids are found only in archaea as a component of polar lipids, more than 25,000 naturally occurring isoprenoid derivatives are known to exist throughout the organisms of the three domains. As isoprenoid biosynthesis is a more ordinary subject than the other aspects of the biosynthesis of archaeal polar lipids, the subject has been more extensively studied and reviewed (9, 93). Therefore, this section is limited to Archaea-specific topics of isoprenoid biosynthesis.

### MVA Pathway from Acetyl-CoA to DMAPP

The isoprenoid biosynthetic pathway is divided into two sections. The first half is the synthesis from acetyl-coenzyme A (CoA) to isopentenyl diphosphate (IPP) and dimethylallyl diphosphate (DMAPP), and the second half is the synthesis of polypropenyl diphosphate from the two C<sub>2</sub> units (IPP and DMAPP). For IPP synthesis, two independent pathways are known: the classical mevalonate (MVA) pathway (Fig. 2) and a more recently discovered MVA-independent (1-deoxy-D-xylulose 5-phosphate [DOXP]) pathway (92). The former is usually found in eucarya and the latter in bacteria, algae, and higher plants. Which pathway is functional in archaea for the formation of the isoprenoid chains of polar lipids is a fundamental question. This was first evidenced by in vivo incorporation experiments of exogenously supplied, isotopically labeled acetate into isoprenoid chains. When <sup>13</sup>CH<sub>3</sub>acetate was incorporated into isoprenoid moieties of polar lipids in Methanospiillum hungatei (28) and Sulfolobus solfataricus (18), <sup>13</sup>C appeared at the positions of methyl carbons (C-3, C-7, C-11, and C-15) and at the carbon positions C-1, C-5, C-9, and C-13 when <sup>13</sup>COOHacetate was fed to the same archaeal cultures. These results are consistent with the isoprenoid biosynthesis pathway from three molecules of acetyl-CoA known in eucarya (Fig. 3). However, Ekiel et al. (29) reported that in Halobacterium cutirubrum, neither <sup>13</sup>COOHacetate nor <sup>13</sup>CH<sub>3</sub>acetate was incorporated into branch-methyl and methine carbons in phytanyl chains; instead, they were derived from lysine. Because <sup>14</sup>Cmevalonic acid was efficiently incorporated into lipids, and carbons at positions other than branch and methine were labeled by acetate, a new route for the formation of MVA from the two molecules of acetate (acetyl-CoA) and lysine was presumed. However, this result has not been adequately evaluated because no specific biochemical reaction was delineated.

It was reported that farnesol inhibits the growth of Halofex volcanii and lipid synthesis (98). The inhibition is attributed to an inhibition of acetate incorporation into lipids, which is recovered by the addition of MVA. Farnesol did not affect the incorporation of MVA. Accordingly, it was concluded that farnesol inhibits the synthesis of MVA from acetate. This phenomenon suggests the existence of a regulatory mechanism of isoprenoid synthesis by farnesol in this organism. It is not known whether farnesol inhibition occurs in other archaea.

An early in vitro work on C<sub>40</sub> carotene synthesis in Halobacterium cutirubrum also showed that isoprenoids are synthesized from MVA via IPP, even though carotenes are not components of polar lipids (57). In vitro assays of enzyme activities of the pathway supported the conclusion led by the in vivo evidence. 3-Hydroxy-3-methyl-glutaryl CoA (HMG-CoA) reductase, a key enzyme of the MVA pathway, was detected, purified, and characterized from Halofex volcanii (6) and Sulfolobus solfataricus (7). The Halofex enzyme was found to be sensitive to lovastatin, an inhibitor of HMG-CoA reductase in mammals. The genes encoding HMG-CoA reductase in Halofex volcanii and Sulfolobus solfataricus were cloned and expressed in Esch-

### Table 1. Nomenclatural changes in archaea discussed in the present text

<table>
<thead>
<tr>
<th>Original name</th>
<th>Strain</th>
<th>Name most recently proposed</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>Caldariella acidophila</td>
<td></td>
<td>Sulfolobus solfataricus</td>
<td>116</td>
</tr>
<tr>
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<td></td>
<td>Halobacterium salinarum</td>
<td>104</td>
</tr>
<tr>
<td>Halobacterium halobium</td>
<td></td>
<td>Halobacterium salinarum</td>
<td>104</td>
</tr>
<tr>
<td>Halobacterium mediterranei</td>
<td></td>
<td>Haloferax mediterranei</td>
<td>102</td>
</tr>
<tr>
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<td></td>
<td>Haloarcula vallismortis</td>
<td>102</td>
</tr>
<tr>
<td>Methanobacterium thermoautotrophicum</td>
<td>ΔH</td>
<td>Methanobacterium thermotaurostrophicus</td>
<td>107</td>
</tr>
<tr>
<td>Methanobacterium thermoautozitrophicum</td>
<td>Marburg</td>
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<td>107</td>
</tr>
<tr>
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<td>SF-4</td>
<td>Methanotrophiobacter wolfii</td>
<td>107</td>
</tr>
<tr>
<td>Methanococcus igneus</td>
<td></td>
<td>Methanococcus igneus</td>
<td>108</td>
</tr>
<tr>
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<td></td>
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</tr>
<tr>
<td>Pseudomonas salinaria</td>
<td></td>
<td>Halobacterium salinarum</td>
<td>32</td>
</tr>
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*E. coli* cells. *Sulfolobus* HMG-CoA reductase showed more than 40% similarity to eucaryal homologs (7). The purified enzymes from both archael species displayed similar kinetic properties to the mammalian enzyme. The HMG-CoA reductases from various eucarya and bacteria were divided into two classes based on amino acid sequences: class I includes the eucaryal enzymes, and class II includes mainly the enzymes from bacteria (8). Archaeal HMG-CoA reductases belong to class I, with the exception of the enzyme from *Archaeoglobus fulgidus*, which is a class II enzyme and is hypothesized to be laterally transferred from bacteria (9).

The seven enzymes and their genes in the MVA pathway from acetyl-CoA to DMAPP are most completely characterized for yeast. When a BLAST search was performed with these sequences as queries, three enzymes had no match in any archael genome (93). Orthologous genes for the four enzymes in the first half of the MVA pathway (acetoacetyl-CoA synthase, HMG-CoA synthase, HMG-CoA reductase, and MVA kinase) were found in the archael genome, while the remaining enzymes were absent.

![MVA pathway](image-url)

**FIG. 2.** MVA pathway for synthesis of isopentenyl diphosphate and dimethylallyl diphosphate. 1, acetyl-CoA acetyltransferase; 2, HMG-CoA synthase; 3, HMG-CoA reductase; 4, MVA kinase; 5, phosphomevalonate kinase; 6, diphosphomevalonate decarboxylase; 7, isopentenyl diphosphate isomerase; 8, hypothetical phosphomevalonate decarboxylase; 9, isopentenyl phosphate kinase. The classical MVA pathway proceeds from reaction 1 through reaction 7 via reactions 5 and 6, while a modified MVA pathway goes through reactions 8 and 9 (33). P and PP in the structural formula are phosphate and pyrophosphate, respectively.
kinase) were detected, while the three orthologs for phosphomevalonate kinase, diposphomevalonate decarboxylase, and IPP isomerase were not. Smit and Mushegian (93), based on an analysis of the sequence motifs of the known enzymes, found candidate enzymes for the missing steps in superfamilies of galactokinase, nucleoside monophosphate kinase, and MutT protein (8-oxo-7,8-dihydro-dGTP pyrophosphohydrolase [62]) in archaeal genomes. They inferred three genes encoding the three enzymes based on linkages, phylogenetic relationships, and sequence similarities. On the other hand, Growchouski et al. (33) have identified isopentenyl phosphate (IP) kinase as an MJ0044 gene product that is one of the isoprene biosynthesis genes of the hyperthermophilic methanoarchaeon *Methanocaldococcus jannaschii*. The presence of IP kinase, in conjunction with the absence of phosphomevalonate kinase and diposphomevalonate decarboxylase, led them to infer that an alternative route for the formation of IPP may be operating in this organism and some related archaea. Their alternative route includes formation of IP from phosphomevalonate by phosphomevalonate decarboxylase and conversion of IP to IPP by IP kinase. The former enzyme is speculative and needs biochemical confirmation. Thus, modification of the MVA pathway is a possible candidate for the IPP synthetic mechanism, at least in seven species of archaea.

Although IPP isomerase is not essential for *E. coli*, in which IPP is synthesized via the DOXP pathway (34), IPP isomerase is essential for organisms with the MVA pathway to form DMAPP, which is the starting allylic C₅ precursor for polyrenyl diphosphate synthesis. IPP isomerase activity, however, could not be detected in any archaea until 2004, when genes from two archaeal species, *Methanothermobacter thermautotrophicus* (2) and *Sulfolobus shibatae* (110), homologous to the *Streptococcus* sp. strain CL190 type 2 IPP isomerase gene *fni* (46), were cloned and expressed in *E. coli* cells. This new type of IPP isomerase requires NAD(P)H and Mg²⁺ and is strictly dependent on flavin mononucleotide without a net redox change (37). The previously known type 1 IPP isomerase does not require these coenzymes. *fni* homologs are found in the whole-genome sequences of many archaea. Thus, it has been verified that in archaea the MVA pathway is responsible for the formation of the two essential C₅ intermediates (IPP and DMAPP) for the biosynthesis of isoprenoids, and no evidence for the DOXP pathway has been obtained.

### Polyprenyl Diphosphate Synthesis

The involvement of polyprenyl diphosphate in polar lipid synthesis was supported by the inhibition of growth and polar lipid synthesis in *Halobacterium cutirubrum* by bacitracin. The inhibitory effect is attributed to its complexing with polyprenyl diphosphates (67).

DMAPP is consecutively condensed with several IPP molecules by prenyltransferase (polyprenyl synthase). The products are geranyl (C₁₀), farnesyl (C₁₅), geranylgeranyl (C₂₀), and farnesylgeranyl (C₂₅) diphosphate (Fig. 4). The products are all in the...
trans form. On the other hand, cis-polyisoprenyl synthase has also been characterized (39); cis-polyisoprenyl diphosphate is not a precursor of membrane polar ether lipids but is possibly a glycosyl carrier precursor. Therefore, we do not discuss cis-prenyl diphosphate synthase in the present paper. The mechanism of each condensing reaction is quite similar; i.e., the allyl diphosphate (DAMPP) serves as an acceptor of IPP. The product of the first condensation is also an allyl diphosphate, which can in turn react with another molecule of IPP to form a product that is a single C₅ unit longer. These enzymes with definite product specificity can synthesize products of shorter chain lengths. Polyisoprenyl synthases are common throughout the three domains of living organisms, i.e., yeast, Archaea, and Bacteria. In Archaea, geranylgeranyl diphosphate (GGPP) synthase was found in Methanobacterium thermoformicum SF-4 (100) (reclassified Methanobacterium wolfeii [107]), Methanobacterium marburgensis (12), Sulfolobus acidocaldarius (85), and Pyrococcus horikoshii OT-3 (64). GGPP synthase also produces farnesyl diphosphate from DMAPP and IPP. The catalytic mechanism of a GGPP synthase from Methanobacterium thermoformicum SF-4 was shown to be an ordered-sequential Bi-Bi mechanism. Potassium ions stimulate the enzyme activity by expediting binding of the substrates (99). Farnesylgeranyl diphosphate synthase was found in Natronobacterium pharaonis (97) (later reclassified Natronomonas pharaonis [45]) and Aeropyrum pernix (101), whose polar lipids contain C₂₅ farnesylgeranyl chains (20, 71). The gene encoding GGPP synthase (idsA) was cloned, identified, and expressed from Methanobacterium marburgensis (11), Sulfolobus acidocaldarius (85, 87), and Pyrococcus horikoshii (64). These enzymes are characterized by aspartate-rich motifs in their sequences, which are commonly found in polyisoprenyl transferases.

As the molecular mechanism for regulation of the chain length of the polyisoprenyl products of GGPP synthase of bacteria and archaea has already been reviewed by Liang et al. (61), and that article discussed the mechanism of chain length determination in Sulfolobus acidocaldarius by Ohnuma et al. (82–84), along with bacteria and eucarya, this subject is no longer dealt with in this review.

A salvage pathway for polyisoprenols was detected in Sulfolobus acidocaldarius, in which geranylgeranoyl and geranylglyceranyl monophosphate were phosphorylated with ATP by separate enzymes (86).

In summary, archaean isoprenoid synthesis proceeds via the classical MVA pathway that is shared with Eucarya, or its modified form; however, the archaean MVA pathway is a mosaic composed of the enzymes common to Archaea and Eucarya, along with enzymes unique to Archaea and Bacteria. In particular, type 2 IPP isomerase and IK kinase, which are the latter enzymes, are remarkably novel. This illustrates that elucidation not only of a metabolic pathway but also of the specifically relevant enzymes is important for obtaining a deeper insight into the biochemistry of lipid metabolism.

FORMATION OF THE G-1-P BACKBONE

In Vivo Evidence

The difference in the stereoconfiguration of the glycerophosphate backbone is one of the most remarkable characteristics that distinguishes archaea from bacteria and eucarya. G-1-P is the enantiomer of bacterial G-3-P. Formation of the G-1-P backbone of polar lipids was investigated in vivo in 1970. Kates et al. (50) first reported radioactively labeled glycerol incorporation into an extremely halophilic archaeon, Halobacterium cutirubrum. When [1(3)-¹⁴C]glycerol and [2-³H]glycerol were incorporated into a glycerol moiety of polar lipids, the ¹⁴C/³H ratio was greatly reduced compared with the ratio of the precursors, while [1(3)-³H]glycerol was incorporated into lipids without a loss of ³H. Kates et al. also detected glycerophosphate dehydrogenase and glycerol kinase activities in this organism, and these were both G-3-P specific (106). They postulated that retention of ³H at the 1 position of glycerol excluded the involvement of dihydroxycetonephosphate (DHAP), which undergoes keto-aldehyde isomerization between DHAP and D-glyceraldehyde-3-phosphate (GAP) by the high level of activity of triose phosphate isomerase. These observations led to the hypothesis that exogenously supplied glycerol was oxidized at the 2 position, forming dihydroxyacetone (DHA), on which one ether linkage is formed at the 1 position. The alkylation of DHA is followed by reoxidation at the 2 position, a second ether bond formation, and phosphorylation. At the reoxidation stage, the stereocconfiguration of the new hydroxyl group is formed so that the first ether bond should be located at the sn-3 position. G-1-P-forming enzymes (glycerophosphate dehydrogenase and glycerol kinase) would not be involved in this pathway. Instead, glycerol dehydrogenase activity was detected in the cell extract of the same organism (4) (the organism called Pseudomonas salinaria at that time [1954] was later renamed Halobacterium cutirubrum and then Halobacterium salinarum [32]). However, the occurrence of glycerol dehydrogenase was species dependent in the genus Halobacterium. While Halobacterium salinarum, Halobacterium cutirubrum, and Halobacterium halobium (these were later reclassified as the same species [104]) have high activities of glycerol dehydrogenase, Halobacterium mediterranei (Halofexar mediterranei [102]) and Halobacterium vallismortis (Haloarcula vallismortis [102]) do not (91). If the speculated pathway is actually operating in halobacterial cells, all of these species must have this enzyme.

Kakinuma et al. (44) conducted similar experiments using [³H]glycerol and nuclear magnetic resonance (NMR) detection. They synthesized position-specific labeled glycerol. ³H of [sn-1-³H]glycerol supplied to a culture of Halobacterium appeared at the sn-3 position of lipid glycerol, while the ²H of the [sn-3-²H]glycerol precursor was incorporated into the sn-1 position of the lipid. This implies that a prochiral glycerol molecule is inverted during lipid biosynthesis. Based on this result, they proposed that an asymmetrical intermediate should be involved in lipid biosynthesis, and the involvement of DHAP instead of DHA was suggested to be the candidate for the asymmetrical intermediate. Kakinuma’s pathway of glycerol incorporation into lipid is as follows: glycerol → G-3-P → DHAP → alkyl DHAP → alkyl G-1-P → dialkyl G-1-P (archaeacidic acid). This model is similar to the Kates model but is different at the phosphorylation step in the pathway. Enzymes that alkylate DHAP are present in mammalian cells (103) but have not been found in any archaea (see below). They included G-3-P but not G-1-P in their proposal, since Kates et al. (106) reported that the glycerophosphate-forming enzyme of the
organism was specific to G-3-P. Although the pathway was thought to be unnatural because of the presence of unnecessary or fruitless oxidation and rereduction of the glycerol moiety at the sn-2 position in the pathway, no evidence against the pathway was presented until Poulter et al. (115) reported a G-1-P-specific ether bond-forming enzyme.

Direct Participation of G-1-P

Poulter et al. (115) investigated in vitro ether bond formation in *Methanothermobacter marburgensis* and found a G-1-P-specific ether bond-forming enzyme (see the next section for details). This suggested that G-1-P was directly involved in the formation of ether lipids. Therefore, direct formation of G-1-P came to be the focus as the next problem. Three reactions were considered to be candidate mechanisms for G-1-P formation (Fig. 5). One was the reduction of GAP at the 1 position (Fig. 5A). Because D-GAP has the same configuration as that of G-1-P at the 2 position, simple reduction of its aldehyde group can form G-1-P. The second possibility for the G-1-P-forming reaction was reduction of DHAP at the 2 position (Fig. 5B). This reaction is similar to the G-3-P dehydrogenase reaction except for the stereospecificity. The last possibility was direct phosphorylation of glycerol at the sn-1 position by ATP (Fig. 5C).

**FIG. 5.** Three possible reactions for the direct formation of G-1-P: A, reduction of D-GAP; B, reduction of DHAP; C, phosphorylation of glycerol (G-1-P forming). Enzyme 1, glycerol kinase (G-3-P forming); enzyme 2, G-3-P dehydrogenase; enzyme 3, triose phosphate isomerase; enzyme 4, G-1-P dehydrogenase. Reaction 5, ether lipid synthesis. FDP, fructose diphosphate.

**Discovery of G-1-P Dehydrogenase (EC 1.1.1.261)**

Nishihara et al. (75) detected glycerophosphate-forming activity from DHAP in a cell-free homogenate of *Methanothermobacter thermautotrophicus*. The homogenate also contained extremely high triose phosphate isomerase activity, which catalyzes interconversion between GAP and DHAP. Therefore, glycerophosphate was apparently formed also from GAP when GAP and NADH were incubated with the unfractionated homogenate. At this stage, the first and second possibilities could not be discriminated. Glycerophosphate formation from GAP was proven to be comprised of combined reactions of triose
phosphate isomerase and glyceraldehyde 3-phosphate dehydrogenase by fractionation of the two enzymes with a DEAE-cellulose column. Because the fractions of cell-free homogenates including no triose phosphate isomerase activity did not exhibit glycerophosphate formation activity from GAP, the possibility of an enzyme catalyzing direct formation of glycerophosphate from GAP was excluded. The activity that catalyzes glycerophosphate formation from DHAP was purified to homogeneity (74). The reaction product from DHAP was confirmed to be G-1-P, and G-1-P but not G-3-P was oxidized in the presence of NAD. NADH acted as a coenzyme, while NADPH was also shown to be active but at a significantly lower level than NADH. Therefore, this enzyme was established as a new enzyme of G-1-P dehydrogenase and has been designated EC 1.1.1.261.

**Interpretation of the In Vivo Phenomena Observed in Halobacterium**

G-1-P-forming activity has been detected in cell extracts of all of the archaeal species so far examined, such as *Methanothermobacter thermautotrophicus*, *Methanosarcina barkeri*, *Halobacterium salinarum*, *Pyrococcus furiosus*, *Pyrococcus sp.* strain KS8-1, and *Thermoplasma acidophilum* strain HO-62 (79, 111). G-3-P was also formed from DHAP by incubation with NADH or NADPH and cell extracts from some of the species (*Methanothermobacter thermautotrophicus*, *Halobacterium salinarum*, *Pyrococcus sp.* strain KS8-1, and *Thermoplasma acidophilum* strain HO-62). G-1-P dehydrogenase activity was also confirmed in recombinant proteins from *Aeropyrum pernix* (36) and *Sulfolobus tokodaiii* (54). G-1-P has never formed from glycerol and ATP, while G-3-P is formed from glycerol and ATP by cell extracts from *Halobacterium*, *Pyrococcus*, and *Thermoplasma*. These are all heterotrophs. Accordingly, the third possibility for G-1-P formation has been excluded. Nishihara et al. (79) discussed the fact that only heterotrophic archaea contain a G-3-P-specific enzyme set (both G-3-P dehydrogenase and G-3-P-forming glycerol kinase), as follows (Fig. 6). When these heterotrophic archaea utilize glycerol as a carbon or energy source, they convert glycerol to G-3-P but not to G-1-P. The produced G-3-P is further metabolized to DHAP by G-3-P dehydrogenase, and DHAP enters the central metabolic pathway after conversion to GAP. In this scenario, G-3-P is used for glycerol catabolism. G-1-P is used for lipid biosynthesis in archaea. DHAP is the crossing point of both pathways. These two pathways hence might confuse the results of the in vivo incorporation experiments performed by Kates et al. and Kakinuma et al. Apparent inversion of the prochiral stereostucture of glycerol should be seen only in the case of the incorporation of exogenously supplied glycerol into lipids in heterotrophic archaea that is not essential for lipid synthesis itself. Although Kates et al. regarded retention of H at the 1 position of glycerol as evidence against the involvement of DHAP because DHAP is in equilibrium with GAP, the absence of any loss of the $^3$H of [1-$^3$H]glycerol during incorporation into lipids might be interpreted by the large equilibrium constant between DHAP and GAP ($K = [DHAP]/[GAP] = 22$ [5]). Because the equilibrium greatly favors DHAP, it is conceivable that the DHAP produced from glycerol via G-3-P is quickly reduced to G-1-P without substantial conversion to GAP. Alternatively, there might be separate pools of DHAP for lipid synthesis and catabolism.

Kakinuma et al. (42) also reported that when D-[6,6-$^2$H]glucose was fed to a *Halobacterium* culture, deuterium appeared at the sn-1 position of lipid glycerol. Because glucose is metabolized via a modified Entner-Doudoroff pathway, carbons at the 4, 5, and 6 positions of glucose are converted to the carbons 1, 2, and 3 of the d-GAP that is isomerized to DHAP. Therefore, this result of Kakinuma is consistent with the in-vivo metabolism of halobacteria.
volvement of G-1-P dehydrogenase proposed by Nishihara et al.

**Phylogenetic Relationships and Molecular Mechanisms of G-1-P Dehydrogenases**

The gene encoding G-1-P dehydrogenase of *Methanothermobacter thermautotrophicus* was cloned, sequenced, named *egsA* (51), and heterologously expressed in *E. coli* (81). The deduced amino acid sequence of G-1-P dehydrogenase from *Methanothermobacter thermautotrophicus* is composed of 347 amino acid residues with a molecular weight of 36,963. Archaeal G-1-P dehydrogenase and bacterial G-3-P dehydrogenase share little sequence similarity, showing that they belong to different enzyme families. A database search detected open reading frames (ORFs) homologous to the *egsA* gene in all 21 species of archaea whose whole genomes had been sequenced up until that time, and it was shown that the archaeal enzyme exhibited sequence similarity to glycerol dehydrogenase, dehydroquinase synthase, and alcohol dehydrogenase (16). A phylogenetic tree of G-1-P dehydrogenases and their homologs was constructed (Fig. 7). Glycerol dehydrogenase, with coordinates available at that time, was shown to be closely related to G-1-P dehydrogenase. Using the structure of glycerol dehydrogenase as the template, Daiyasu et al. (16) built a model structure of G-1-P dehydrogenase that predicts the following structure and function characteristics of the enzyme: (i) the chirality of the product, (ii) the requirement of the Zn$^{2+}$ ion for the enzyme reaction, (iii) the transfer of pro-R hydrogen of NADH during the enzyme reaction, (iv) the putative active site and the reaction mechanism, and (v) that G-1-P dehydrogenase does not share an evolutionary origin with G-3-P dehydrogenase from bacteria. The pro-R hydrogen transfer and Zn$^{2+}$ requirement were experimentally verified (35, 55). It is known that G-3-P dehydrogenase transfers the pro-S hydrogen of NADH (23). Crystallographic data showed that *pro-R* stereospecific enzymes and *pro-S* stereospecific enzymes bind the nicotinamide ring of the coenzyme at the opposite orientation, and this is apparently the basis for the enzyme stereospecificity (112). Therefore, it is assumed that G-1-P dehydrogenase (*pro-R* type) and G-3-P dehydrogenase (*pro-S* type) have symmetrical ternary structures, at least in the coenzyme-binding sites. G-1-P dehydrogenase in *Aeropyrum pernix* has been kinetically studied (36). The enzyme uses NADH or NADPH as a coenzyme in DHAP reduction with preference for NADPH but does not use NADP in G-1-P oxidation. This factor, along with the far lower $K_m$ value for DHAP than for G-1-P (74), suggests that G-1-P dehydrogenase is really functioning in the direction of G-1-P formation from DHAP. A kinetic analysis revealed that the catalytic reaction by the enzyme follows an ordered Bi-Bi mechanism (36).

Thus, G-1-P dehydrogenase has been established at the enzyme and genetic levels as a key enzyme responsible for the formation of the enantiomeric glycerophosphate backbone structure of archaeal phospholipids.

**G-1-P Formation in Sulfolobus**

An example that cannot be explained by the G-1-P pathway is the case of *Sulfolobus*. [U-$^{13}$C,1(3)-$^3$H]glycerol and [U-$^{13}$C,2-$^3$H]glycerol were incorporated into lipid glycerol without any loss of $^3$H in vivo labeling experiments in *Calderilla acidophilus* (Sulfolobus solfataricus) (116) (21). Kakinuma et al. confirmed this result by using stereospecifically labeled glycerol with $^3$H and reported no inversion of the glycerol moiety of lipids during biosynthesis in *Sulfolobus acidocaldarius* (41). One possible explanation for these results may be direct phosphorylation of glycerol by an unknown G-1-P-forming glycerol kinase. Only one study, presented orally at a meeting, has thus far reported the product of glycerol kinase in *S. acidocaldarius* to be G-3-P (81a). Although the phenomena described by De Rosa et al. and Kakinuma et al. remain to be explained, the universality of the presence of G-1-P dehydrogenase was verified by the detection of G-1-P dehydrogenase in a species of *Sulfolobus* (S. tokodaiii) (54).

**ETHER BOND FORMATION**

The pathways for biosynthesis of the ether-type and ester-type phospholipids from archaea and bacteria, respectively, are depicted in Fig. 8. **First Ether Bond Formation (G-1-P: GGPP Geranylgeranyltransferase; GGPP Synthase; EC 2.5.1.41)**

Ether lipid biosynthesis was studied by in vivo incorporation of several lipid components into *Methanospirillum hungatei* cells (90). Archaeol (2,3-di-O-phytanyl-sn-glycerol) and caldarchoeol (2,3,2,3′-di-O-bisphytylanyl-di-sn-glycerol) were incorporated into mainly the nonpolar lipid fraction, and no interconversion between them was found. Among the C$_{20}$ polyenyls, fully unsaturated geranylgeraniol was most efficiently incorporated into polar lipids in intact cells of the methanogenic archaeon. Monounsaturated phytol was poorly incorporated, and fully saturated phytanol was not incorporated. The results suggested that an ether bond was formed from unsaturated prenyl precursors (90). This was confirmed by an in vitro enzymatic experiment with sonicates of *Methanothermobacter marburgensis* cells (115). Ether bond formation is catalyzed by two prenyl transferases: one is responsible for formation of the first ether bond between the sn-3 hydroxyl group of G-1-P and GGPP (GGPP synthase), and the other catalyzes the formation of the second ether bond at the sn-2 position to form di-O-geranylgeranyl-G-1-P (DGGGP, or unsaturated archaeatic acid) (DGGGP synthase) (114). The first enzyme was found in sonic extracts of *Methanothermobacter marburgensis* cells and *Halobacterium halobium* cells (113). The enzyme showed 30 to 40 times higher activity to G-1-P than to G-3-P and did not react with DHAP. This evidence directly contradicts Kakinuma’s hypothesis. GGPP synthase exhibited the same order of specificity of geranylgeranyl-, phytanyl-, and phytanyl diphosphate as the incorporation efficiencies of these prenyl alcohols exhibited in the in vivo incorporation experiments (115). Farnesyl diphosphate could not serve as a substrate for GGPP synthase (114). GGPP synthase is a soluble enzyme, while the second ether bond-forming enzyme is associated with the membrane fraction (114). GGPP synthase was first purified to homogeneity.
FIG. 7. Phylogenetic tree of G-1-P dehydrogenase and its homologs. The sequence is indicated by the source name of the sequence database (sp, SwissProt; pir, PIR; gb, GenBank; pdb, PDB) and the identification code. G1PDH, sn-glycerol-1-phosphate dehydrogenase; GDH, glycerol dehydrogenase; DHQS, dehydroquinate synthase; ALDH, alcohol dehydrogenase type IV. (Reprinted from reference 16 by permission of Oxford University Press.)
from the Methanothermobacter marburgensis cytosolic fraction, and its properties were described (13). The genes encoding GGGP synthase from Methanothermobacter marburgensis (94) and Thermoplasma acidophilum (72) were cloned and expressed in E. coli cells. Each recombinant enzyme was purified and characterized. Homologs of the Methanothermobacter marburgensis gene were found in Methanothermobacter thermautotrophicus \( \Delta H \), Methanocaldococcus jannaschii, Pyrococ-

**Archaebacteria**

FIG. 8. Possible biosynthetic pathway for phospholipids in archaebacteria compared with their bacterial counterpart. Enzymes confirmed by in vitro experiments are as follows: 1, G-1-P dehydrogenase; 2, GGGP synthase; 3, DGGGP synthase; 4, CDP-archaeol synthase; 5, archaetidylserine synthase. Reactions and enzymes 6 to 9 are indicated by in vivo experiments or database searches of the relevant genes. 6, archaetidylserine decarboxylase; 7, archaetidylinositol synthase; 8, archaetidylglycerophosphate synthase; 9, archaetidylglycerophosphate phosphatase. The established enzymes for phospholipid biosynthesis in bacteria are as follows; 1', G-3-P dehydrogenase; 2', lysophosphatidic acid synthase; 3', phosphatidic acid synthase; 4', CDP-diacylglycerol synthase; 5', phosphatidylserine synthase; 6', phosphatidylserine decarboxylase; 7', phosphatidylinositol synthase; 8', phosphatidylglycerophosphate synthase; 9', phosphatidylglycerophosphate phosphatase. P and PP in the structural formula are phosphate and pyrophosphate, respectively.
**Second Ether Bond Formation (GGGP; GGPP Geranylgeranyltransferase; DGGGP Synthase; EC 2.5.1.42)**

The second ether bond-forming enzyme was studied for two species of archaea. Digeranylgeranyl G-1-P (unsaturated arachaeatic acid, or DGGGP) synthase activity was found in the membrane fraction of *Methanothermobacter marburgensis* (114). It was also specific to the G-1-P-derived substrate, monogera-

nylgeranyl G-1-P, but did not react with 1-monogeranylgeranyl-

G-3-P (GGGP + GGPP → GGGP [archaeatic acid] + PP). Hemmi et al. (38) found that *Sulfolobus solfataricus* DGGGP synthase was one of the three enzymes that are members of the UbA prenyl transferase family, which, excluding DGGGP synthase, transfers prenyl groups to hydrophobic ring structures such as quinones, hemes, chlorophylls, vitamin E, or shikonin. One of the encoding genes, SS00583, was cloned and expressed in *E. coli*, and the recombinant protein was confirmed to have DGGGP synthase activity. They discussed the phylogenetic relationship of DGGGP synthase with the other UbA homologs, such as the side chain production enzymes of respiratory quinones and hemes. These ether bond-forming enzymes, in addition to G-1-P dehydrogenase, help establish and maintain the G-1-P structure in polar lipids in archaea by their substrate specificity.

**ATTACHMENT OF POLAR HEAD GROUPS**

**In Vivo Pulse-Labeling Experiments**

Kates and coworkers explored the biosynthetic pathways of polar lipids (phospholipid, glycolipid) in *Halobacterium cutirubrum* in vivo (66). Three kinds of radioactive precursors ([14C]MVA, [14C]glycerol, and 32P) were separately supplied to the culture of the organism for a short time (1 to 60 min). The labeled lipids were separated by thin-layer chromatography (TLC) and identified by various chemical degradation procedures (acid methanolysis of phosphodiester linkages and glycosyl linkages, alkaline hydrolysis of phosphodiester linkages, and Vitride hydrogenolysis [reductive degradative of allyl ether compounds with sodium dihydrobis(2-methoxyethoxy)-aluminate]) and mass spectrometry. The biosynthetic relationship of the intermediates was inferred from the time course of labeling of each lipid. The intermediates of the short-term pulse labeling were classified into two types: “early” and “later.” The early intermediates represent short-term precursors that appear within the first few minutes of biosynthesis and are converted to the later intermediates. The latter are composed of the same polar head groups as the final polar lipid products but with acid-labile allyl ether hydrocarbon chains. These later intermediates were termed “pre-PGP,” “pre-PG,” “pre-PGS,” and “pre-S-TGD.” PG, PG, PGS, are saturated archaeaticlyglycerophosphate, archaeaticlyglycerol, and archaeati-
dyglycerolate, respectively. S-TGD is sulfated triglycerolyl-

chaol. The early intermediates probably include unsaturated prenyl phosphate, unsaturated archaeatic acid, unsaturated CDP-archaol, and unidentified intermediates. On the basis of these results, the authors presented the following tentative pathway for the biosynthesis of the major polar lipids in *Halobacterium cutirubrum*. Glycerol or dihydroxyacetone is alkylated with phytol diphasphate or GGPP to give di-isoprenylglycerol or the substi-
tuted glycerol derivative, which is then phosphorylated to “pre-PA.” This early intermediate appears to be converted to “pre-PGP” via putative unsaturated CDP-archaol. Dephosphorylation of “pre-PGP” and then the sulfating of it give “pre-PG” and “pre-PGS.” Hydrogenation of the “pre-” intermediate gives the saturated final products. Dephosphorylation of “pre-PA” enters the biosynthetic pathway for glycolipids. It is significant that Moldoveanu et al. (66) pointed out the involvement of unsaturated intermediates in polar lipid synthesis in archaea before Poulter et al. (115) showed the results of in vitro experiments that the first ether bond-containing intermediate was unsaturated GGGP.

**CDP-Archaol Synthase (CTP:2,3-di-O-Geranyloglyceral-1-Phosphate Cytidyltransferase)**

The above results by Kates et al. (66) outlined the biosynthetic route of polar lipids in archaea, although some parts of the proposed pathway were not confirmed by subsequent in vitro enzymatic investigations. In vitro experiments to clarify ether polar lipid biosynthesis reactions were performed mainly with cell-free homogenates of *Methanothermobacter marburgensis* (ether bond formation) or *Methanothermobacter therm- autotrophicus* ΔH (polar lipid synthesis). As described above, two ether bonds form on the G-1-P molecule but not on DHA or glycerol. The prenyl donor was GGPP rather than phytol.
diphosphate. The first diether-type phospholipid intermediate was found to be digeranylgeranyl G-1-P (unsaturated archaetidic acid). This was designated for spot 11, which is one of the “early” intermediates cited by Moldoveanu and Kates (66). CDP-archaeol was found to be involved in the polar lipid biosynthetic pathway of archaea by Morii and Koga (70). This was nominated as the entity of another early intermediate, spot 4. In contrast to these intermediates and reactions, the identity of the nonphosphorylated early intermediate spot 12 has yet to be delineated by in vitro experiments. Although these differences in polar lipid biosynthetic mechanisms can be attributed to the difference of species (an extreme halophile and a methanogen), it is reasonable to consider that a general biosynthetic mechanism is shared by both archaeal species, since the enzyme activities and the genes of G-1-P dehydrogenase, ether bond-forming enzymes, and acid-labile unsaturated intermediates are all held in common.

Morii and Koga (70) detected, in the Methanothermobacter thermautotrophicus cell membrane fraction, an activity that catalyzes the synthesis of CDP-archaeol from unsaturated archaetidic acid and CTP (CTP,2,3-di-O-geranyleranyl-sn-glycerol-1-phosphate cytidylyltransferase) (DGGGP [archaetidic acid] + CTP → CDP-archaeol + PP). The product of the reaction was identified as CDP-digeranylgeranylcylerol by chromatographic mobility, chemical analysis, acid lability, and mass spectrometry. A tiny amount of radioactive CDP-archaeol was detected in growing cells, which was briefly labeled with [32P]CTP and the membrane fraction of Methanothermobacter thermautotrophicus at 55°C for 2 h. After the reaction, chloroform-extractable [3H]H was counted.

**Table 2. Substrate specificity of CDP-archaeol synthase from Methanothermobacter thermautotrophicus**

<table>
<thead>
<tr>
<th>Substrate</th>
<th>Hydrocarbon</th>
<th>No. of double bonds/side chain</th>
<th>Linkage</th>
<th>GP backbone</th>
<th>CDP-archaeol synthase activity (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>2,3-GG-GP ether</td>
<td>Geranylgeranyl</td>
<td>4</td>
<td>Ether</td>
<td>G-1-P</td>
<td>100</td>
</tr>
<tr>
<td>1,2-GG-GP ether</td>
<td>Geranylgeranyl</td>
<td>4</td>
<td>Ether</td>
<td>G-3-P</td>
<td>94</td>
</tr>
<tr>
<td>2,3-Phyta-GP ether</td>
<td>Phytyl</td>
<td>0</td>
<td>Ether</td>
<td>G-1-P</td>
<td>3</td>
</tr>
<tr>
<td>2,3-Phyta-GP ether</td>
<td>Phytyl</td>
<td>1</td>
<td>Ether</td>
<td>G-1-P</td>
<td>0</td>
</tr>
<tr>
<td>2,3-Ole-GP ether</td>
<td>Oleyl</td>
<td>1</td>
<td>Ether</td>
<td>G-1-P</td>
<td>0</td>
</tr>
<tr>
<td>2,3-GG-GP ester</td>
<td>Geranylgeranoyl</td>
<td>4</td>
<td>Ester</td>
<td>G-1-P</td>
<td>122</td>
</tr>
<tr>
<td>1,2-GG-GP ester</td>
<td>Geranylgeranoyl</td>
<td>4</td>
<td>Ester</td>
<td>G-3-P</td>
<td>50</td>
</tr>
<tr>
<td>rac-GG-GP ester</td>
<td>Geranylgeranoyl</td>
<td>4</td>
<td>Ester</td>
<td>rac-GP</td>
<td>81</td>
</tr>
<tr>
<td>2,3-Ole-GP ester</td>
<td>Oleoyl</td>
<td>1</td>
<td>Ester</td>
<td>G-1-P</td>
<td>13</td>
</tr>
<tr>
<td>rac-Ole-GP ester</td>
<td>Oleoyl</td>
<td>1</td>
<td>Ester</td>
<td>rac-GP</td>
<td>3</td>
</tr>
</tbody>
</table>

See reference 70. Each substrate (2,3-di-O-geranyleranyl-sn-glycerol-1-phosphate) analog synthesized was incubated with [5-3H]CTP and the membrane fraction of Methanothermobacter thermautotrophicus at 55°C for 2 h. After the reaction, chloroform-extractable [3H]H was counted.

The biosynthetic pathway of polar lipids in archaea has now been confirmed in vitro from DHAP to archaetidylserine. The sequence of reactions in the pathway resembles the bacterial phospholipid synthesis pathway except for the stereostructure of glycerophosphate, ether bonds, and isoprenoid chains. Both pathways consist of the reduction of DHAP, attachment of two ral chains to glycerophosphate, activation by CTP, and replacement of CMP by various polar groups, in that order.
ically by in vivo pulse-chase experiments with 32Pi, which was not yet been obtained. In vitro enzymatic evidence has not yet been obtained. The conditions for the enzymes might be different. The results suggested that archaetidylethanolamine was synthesized from archaetidylserine by decarboxylation. It was suggested that an ORF encodes archaetidylserine decarboxylase from the data showing sequence similarity with phosphatidylserine decarboxylase and the conserved adjacent location of and the order of the ORF and archaetidylserine synthase in the genome (17). Probable archaetidylserine decarboxylase is expected to transform archaetidylserine to archaetidylethanolamine: archaetidylserine → archaetidylethanolamine + CO₂.

If archaetidylserine decarboxylase is included in the cell-free homogenate and archaetidylserine has been formed from CDP-archaol and L-serine during archaetidylserine synthase reaction, it would be expected that a portion of the archaetidylserine formed might be converted to archaetidylethanolamine in the reaction mixture. When the archaetidylserine synthase reaction was carried out using cell-free homogenates of *Methanothermobacter thermautotrophicus* with proper substrates under optimal conditions, archaetidylserine was readily synthesized but no trace of archaetidylethanolamine was detected in the reaction mixture after the archaetidylserine synthase reaction was completed (H. Morii, unpublished results). The fact that no archaetidylethanolamine was formed suggests that the reaction conditions for the enzymes might be different. The conditions for the archaetidylethanolamine synthase reaction are not known at present.

Serine, ethanolamine, glycerol, and myo-inositol as the polar head groups of phospholipids are shared by *Archaebacteria*, *Bacteria*, and *Eucarya*, in contrast to other structural aspects of phospholipids, such as ether bonds, isoprenoid side chains, and the G-1-P backbone structure. In accordance with the distinctive structural traits of lipids, G-1-P-forming enzymes and ether bond-forming enzymes are specific to *Archaebacteria*, whereas polar head group-attaching enzymes are common to organisms throughout the three domains. This is a hybrid nature in the lipid synthetic pathway; we discuss below what this implies.

Although choline phospholipid has been reported to be present in a species of methanogenic archaeon (*Methanopyrus kandleri*), no homolog for a choline phospholipid-synthesizing enzyme was detected in a database search (17). Therefore, at present no means to discuss choline phospholipid biosynthesis in archaea is available. The unrelatedness of the choline phospholipid-synthesizing enzymes of archaea and bacteria may reflect the deepest branching of *M. kandleri* in phylogeny.

### Homology Search for the CDPAldcohol Phosphatidyltransferase Family

Although only the archaetidylserine synthase from *Methanothermobacter thermautotrophicus* has been biochemically characterized among the archaenae polar lipid-synthesizing enzymes and other enzymes have not been examined, Daiyasu et al. (17) tried a bioinformatics approaches to obtaining more information on polar lipid biosynthesis in archaea. A first search for CDP-diacylglycerol synthase homologs in archaea did not yield significant hits (Y. Koga, unpublished results). From a database search of 22 archaeal species whose entire genomes had been sequenced, many archaenal hypothetical proteins were found to display sequence similarity to members of the CDPAldcohol phosphatidyltransferase family, including phosphatidyleserine synthase, phosphatidylglycerol synthase, and phosphatidylinositol synthase derived from bacteria and eucarya (Fig. 9 and 10) (17). These enzymes may catalyze the following reactions: CDP-archaol + L-serine → archaetidylserine + CMP; CDP-archaol + G-1-P → archaetidylglycerophosphate + CMP; and CDP-archaol + myo-inositol → archaetidylmyo-inositol + CMP. These archaenal proteins were classified into two groups based on sequence similarity. The first group included archaetidylserine synthase from *Methanothermobacter thermautotrophicus* and members that were closely related to phosphatidylserine synthase (Fig. 9). Most of the archaeal species that had these proteins contained serine phospholipid (52). These proteins were thus suggested to be archaetidylserine synthase. The second group of archaenal hypothetical proteins was related to phosphatidylglycerophosphate synthase and phosphatidylinositol synthase (Fig. 10). By constructing a phylogenetic tree of these proteins and considering the distribution of glycerol phospholipid and inositol phospholipid (52), these were divided into two clusters, one of which corresponds to archaetidylglycerophosphate synthase and the other to archaetidylinositol synthase (Fig. 10). This widespread distribution of genes for possible archaetidylglycerophosphate synthase and archaetidylinositol synthase suggests that archaenal glycerol phospholipid and inositol phospholipid would probably be synthesized by the reactions depicted in the above equations and in Fig. 8 and catalyzed by these enzymes, although in vitro enzymatic evidence has not yet been obtained.

### Archaetidylethanolamine Synthesis

Ethanolamine phospholipid biosynthesis was studied kinetically by in vivo pulse-chase experiments with 32P, which was reviewed previously (53). The results suggested that archaetidylethanolamine was synthesized from archaetidylserine by decarboxylation. It was suggested that an ORF encodes archaetidylserine decarboxylase from the data showing sequence similarity with phosphatidylserine decarboxylase and the conserved adjacent location of and the order of the ORF and archaetidylserine synthase in the genome (17). Probable archaetidylserine decarboxylase is expected to transform archaetidylserine to archaetidylethanolamine: archaetidylserine → archaetidylethanolamine + CO₂.

### Table 3. Substrate specificity of archaetidylserine synthase from *Methanothermobacter thermautotrophicus*

<table>
<thead>
<tr>
<th>Substrateb</th>
<th>Hydrocarbon</th>
<th>No. of double bonds/side chain</th>
<th>Linkage</th>
<th>GP backbone</th>
<th>Relative CDP-archaol synthase activity (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>CDP-2,3-GG-Gro ether</td>
<td>Geranylgeranyl</td>
<td>4</td>
<td>Ether</td>
<td>G-1-P</td>
<td>100</td>
</tr>
<tr>
<td>CDP-1,2-GG-Gro ether</td>
<td>Geranylgeranyl</td>
<td>4</td>
<td>Ether</td>
<td>G-3-P</td>
<td>149</td>
</tr>
<tr>
<td>CDP-2,3-Phyta-Gro ether</td>
<td>Phytany</td>
<td>0</td>
<td>Ether</td>
<td>G-1-P</td>
<td>86</td>
</tr>
<tr>
<td>CDP-1,2-Ole-Gro ether</td>
<td>Oleyl</td>
<td>1</td>
<td>Ether</td>
<td>G-3-P</td>
<td>64</td>
</tr>
<tr>
<td>CDP-2,3-Ole-Gro ester</td>
<td>Oleyl</td>
<td>1</td>
<td>Ester</td>
<td>G-1-P</td>
<td>280</td>
</tr>
<tr>
<td>CDP-1,2-acyl-Gro ester</td>
<td>Fatty acyl</td>
<td></td>
<td>Ester</td>
<td>G-3-P</td>
<td>199</td>
</tr>
</tbody>
</table>

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*a See reference 69. Each substrate (CDP-2,3-di-O-geranylgeranyl-α-glycerol or CDP-archaol) analog synthesized was incubated with [3-3H]α-serine and cell-free homogenate of *Methanothermobacter thermautotrophicus* at 60°C for 10 min. After the reaction, chloroform-extractable 3H was counted.

b Abbreviations: GG, geranylgeranyl; Gro, glycerol; Phyta, phytanyl; Ole, oleoyl; acyl, mixed fatty acyl.
Although no biochemical information on archaeal glycerol 3-phosphate synthase and archaealdiglycerol 3-phosphate synthase has been made known to date, archaealglycerol 3-phosphate synthase has been putatively synthesized from CDP-archaeol and myo-inositol, because potential archaealglycerol 3-phosphate synthase belongs to the same enzyme family (the CDP-alcohol phosphatidyglycerol transferase family, as described above). One of the substrates for this enzyme reaction in eucarya is myo-inositol, which is generated from 1L-myoinositol-1-phosphate by dephosphorylation. 1L-myoinositol-1-phosphate is a precursor of di-myoinositol-monophosphate, which is a compatible solute of certain archaeal species, including Archaeoglobus fulgid us, Pyrococcus woesei, Pyrococcus furiosus, and Methanococcus
cus igneus (later reclassified *Methanotorris igneus* [108]). Chen et al. (15) characterized 1L-myoinositol-1-phosphate synthase as the product of the *ips* gene of *Archaeoglobus fulgidus* expressed in *E. coli*. Even though it is an enzyme reported to be involved in the compatible solute biosynthesis, it is most likely that the enzyme reaction product can also be the substrate of arcaetidylinositol synthase. The enzyme catalyzes conversion of glucose-6-phosphate to 1L-myoinositol-1-phosphate in the presence of NAD (D-glucose-6-P + NAD → 1L-myoinositol-1-P + NAD). The stereochemistry of the product was identified as 1L-myoinositol-1-phosphate by FIG. 10. Phylogenetic tree of arcaetidylinositol synthase and arcaetidylglycerol synthase constructed by the maximum-likelihood method. The sequence is indicated by the source name and GI number from the National Center for Biotechnology Information. The presence or absence of arcaetidylinositol (AI) or arcaetidyglycerol (AG)/arcaetidyglycerophosphate (AGP) in each species is indicated by a symbol, “+” or “−,” after the source name and GI number. The symbol “?” indicates that the lipid composition has not been reported for the species. For the characters “A,” “B,” “C,” and “X,” see reference 17. PIS, phosphatidylinositol synthase; PGS, phosphatidyglycerol synthase; AIS, arcaetidylinositol synthase; AGS, arcaetidyglycerol synthase. (Reprinted from reference 17 by permission of the publisher.)
A phylogenetic tree of 1L-myoinositol-1-phosphate synthase was constructed, and the evolutionary relationships were discussed (1). The three-dimensional structure of the Archaeoglobus enzyme was analyzed crystallographically, on the basis of which the reaction mechanism was studied in detail (95). A conversion activity of glucose-6-phosphate to inositol phosphate (1L-myoinositol-1-P → myoinositol + P) was found by the NaIO₄ oxidation-P₂ release method (3) in crude cell homogenates of Methanothermobacter thermoautotrophicus (H. Morii and Y. Koga, unpublished results). The stereostructure of the product was identified as 1L-myoinositol-1-phosphate by use of chiral column–gas-liquid chromatography–mass spectrometry. Because the organism does not contain di-myoinositol-monophosphate as a compatible solute, the 1L-myoinositol-1-phosphate produced would probably be the precursor of archaetidylinositol synthase in this methanocarchaeon.

If archaetidylinositol is synthesized from CDP-archaeol and myo-inositol, 1L-myoinositol-1-phosphate should be dephosphorylated. The 1L-myoinositol-1-phosphate phosphatase gene has been cloned as the subB gene from the Archaeoglobus fulgidus genome and expressed heterologously (14). Based on the above results, archaetidylinositol would be synthesized from CDP-archaeol and myo-inositol, which is a reaction analogous to phosphatidylinositol synthesis.

Hydrogenation of Unsaturated Intermediates

The intermediates thus far identified for phospholipid biosynthesis in archaea are all unsaturated or geranylgeranyl chain-containing phospholipids. On the other hand, the final products or membrane phospholipids in most archaea contain fully saturated chains. Therefore, hydrogenation or saturation reactions must be operating somewhere along the steps leading these intermediates to final products.

Nishimura and Eguchi (80) have purified and characterized an enzyme in Thermoplasma acidophilum that catalyzes reduction (hydrogenation) of the geranylgeranyl chains of unsaturated arachadiacid. Because NADH was required for the reduction and flavin adenine dinucleotide (FAD) increased the activity, NADH would act as a hydrogen donor via FAD. The N-terminal amino acid sequence was determined and the encoding gene was cloned. Although the activity was detected in only Thermoplasma acidophilum cells, homologous genes were found in several archaeal cells. The enzyme also catalyzed the reduction of unsaturated arachidylglycerophosphate, unsaturated arachidylglycerol, and unsaturated arachidylethanolamine. If unsaturated arachidylglycerol were saturated, it could not react with CDP-archaeol synthase, because CDP-archaeol synthase reacts preferentially with unsaturated arachidic acid, as described above, supposing that Thermoplasma acidophilum and Methanothermobacter thermoautotrophicus synthesize their phospholipids by the same biosynthetic pathway. Saturation of the geranylgeranyl chains would therefore take place after the formation of CDP-archaeol. The real substrate of the new reductase might possibly be phospholipids with polar head groups. A gene (afl0464) of Archaeoglobus fulgidus encoding the homologous enzyme has been expressed in E. coli cells, and the enzyme has been purified (71a).

It was not possible to determine the exact step at which saturation occurs, as unsaturated and saturated CDP-archaeols are of almost comparable activity to archaetidylyserine synthase. However, unsaturated CDP-archaeol and unsaturated archaetidylyserine were detected in intact cells of Methanothermobacter thermoautotrophicus, suggesting that the saturation reaction may take place somewhere after attachment of the polar head groups.

Glycolipid Synthesis

Morii and Koga (unpublished data) have also studied the in vitro biosynthesis of gentiobiosyl (β-D-glucoyl-(1→6)-β-D-glucoyl) archaeol. The gentiobiosyl moiety is the only glycosyl group of glycolipids and phosphoglycolipids of Methanothermobacter thermoautotrophicus (76). Cell-free homogenates contained activities that catalyze the transfer of glucose from UDP-glucose to saturated archaeol and the transfer of another glucose unit to monoglucosylarchaeol from UDP-glucose to generate monoglucosylarchaeol (monoglucosylarchaeol synthase) (archaeol + UDP-glucose → monoglucosylarchaeol + UDP) and diglucosylarchaeol (diguclcosylarchaeol synthase) (monoglucosylarchaeol + UDP-glucose → diglucosylarchaeol + UDP). Both activities are loosely associated with membranes, monoglucosylarchaeol synthase being more loosely associated. The two activities are, although not yet separated, distinguished by the requirement of the Mg²⁺ ion for activity and cellular localization. Both activities strictly require archaetidylinositol, similar to Streptococcus pneumoniae diglycosyl diacylglycerol synthase, which requires phosphatidylglycerol or cardiolipin (24). Monoglucosylarchaeol synthase is also active to caldarchaeol and caldarchaetidylinositol (both are tetraether-type lipids), as well as to 2,3-diphytanyl-sn-glycerol (saturated archaeol). 2,3-Digeranylgeranyl-sn-glycerol (unsaturated archaeol) was 30% active compared to saturated archaeol. It is not clear whether the natural substrate of monoglucosylarchaeol synthase is saturated or unsaturated archaeol, since the natural source of saturated or unsaturated archaeol is unknown. The mechanism of biosynthesis of glycolipid with two same hexose units are classified into two cases: in the first, one enzyme catalyzes the transfer of both the first and second (or even third or more) sugar units, and in the second, individual specific enzymes are responsible for the formation of monoglucosyl lipid and diglycosyl lipid. In the case of Methanothermobacter thermoautotrophicus, the latter type is likely as seen in Achaeolepsma (24). The former example is known in Bacillus subtilis gentiobiosyl diacylglycerol synthase (40).

Compared with the glycolipids of Methanothermobacter thermoautotrophicus, which are composed of a rather simple glycosyl moiety (with only glucose as a hexose unit), the glycolipids of thermoacidophilic and halophilic archaea are generally multifarious. The biosynthesis of each glycolipid from those archaea is not understood at all.

BIOSYNTHESIS OF TETRAETHER POLAR LIPIDS

The formation of tetraether lipids is one of the most challenging and interesting problems in all of biochemistry. The ultimate reaction of tetraether lipid synthesis is C-C bond formation between the two methyl termini of phytanyl chains or their precursor chains. Since this is an extremely unusual reaction, never seen previously in biochemistry, no one pres-
ently has a clue regarding how to make it clear in an in vitro reaction. Only the results from in vivo incorporation experiments are currently available. Four kinds of in vivo experiments for the study of tetraether lipid synthesis have been reported.

**Simple Kinetic Studies between Diether and Tetraether (Polar) Lipids**

When *Thermoplasma* cells were labeled with [14C]MVA for a short period, radioactivity was first incorporated into archaeol core lipid, and incorporation of radioactivity into caldarchaeol core lipid increased after the time point at which the labeling of archaeol reached its maximum (59). A pulse-chase experiment revealed a precursor-product relationship between the archaeol and caldarchaeol cores. Langworthy proposed from these results that C_{40} phytanyl diethers might serve as the immediate precursor of C_{40} tetraether biosynthesis via direct condensation of two C_{20} diethers in the membrane (59). Since the author prepared archaeol from diether-type polar lipids, the discussion implies that diether-type polar lipids are the precursors of tetraether-type polar lipid formation but not that the precursor is the bare archaeol core lipid itself. As described above, when archaeol and caldarchaeol were incorporated into the mainly nonpolar lipid fraction of *Methanospirillum hungatei* cells, no interconversion between them took place (90). This suggests that tetraether lipid is not synthesized by condensation of two molecules of diether core lipid (archaeol) without the polar head groups.

A similar type of kinetic study of P_{i} incorporation instead of MVA in a methanogen species was conducted. As this has been discussed in a previous review (53), we summarize it here briefly. Incorporation of P_{i} into phospholipids in growing *Methanothermobacter thermoautotrophicus* cells suggested a precursor-product relationship between diether polar lipids and tetraether polar lipids which had already been substituted by phosphoric esters and glycosyl moieties (76). The previous review (53) discussed other lines of evidence for the synthesis of tetraether polar lipids from diether polar lipids, such as the structural regularity of polar lipids in *Methanothermobacter thermoautotrophicus* (76) and *Methanospirillum hungatei* (58), as well as the asymmetric and similar distribution of diether and tetraether polar lipids in the cytoplasmic membranes of *Methanothermobacter thermoautotrophicus* (68).

**Inhibition of Tetraether Lipid Synthesis**

A recent significant advance in tetraether lipid biosynthesis has been achieved in pulse-chase experiments using an inhibitor (terbinafine) of tetraether lipid formation (56, 73) (Fig. 11). Terbinafine is originally a squalene epoxidase inhibitor. Although it is not known why a squalene epoxidase inhibitor inhibits tetraether lipid synthesis, it in fact does inhibit the formation of tetraether lipids, and a diether polar lipid precursor accumulates as a result. After removal of the inhibitor, the precursor was converted to tetraether polar lipids. A time course of appearing and disappearing radioactivity after chasing in each intermediate lipid spot on a TLC plate established the biosynthetic relationships of the intermediates. The structures of the accumulated intermediates were analyzed. As a result, the following results were obtained. The diether phospholipid which accumulated in the presence of terbinafine is saturated archaetidylglycerol, which is converted to bisarchaetidylglycerol (caldarchaeol with two phosphoglycerols on both of the hydroxyl groups) after removal of the inhibitor. One of the two phosphoglycerol groups on the caldarchaeol core of bisarchaetidylglycerol is removed to give caldarchaetidylglycerol, which is then glycosylated to form glycosyl caldarchaetidylglycerol (the so-called “main polar lipid” of *Thermoplasma acidophilum* [96]). It is most notable that the diether precursor of tetraether lipid formation has saturated isoprenoid chains. However, even though the reaction sequence for tetraether lipid biosynthesis has been outlined by this work, the mechanism of C-C bond formation remains unknown. It is not easy to conceive what sort of reaction would lead to the establishment of a C-C bond between two highly unreactive saturated methyl groups. However, an unpublished result by H. Morii and Y. Koga (as described above) that tetraether-type lipids (caldarchaeol and caldarchaeidyl-myko-inositol) were glucosylated with UDP-glucose in the presence of *Methanothermobacter thermoautotrophicus* cell extracts is consistent with the results of the terbinafine inhibition experiments with *Thermoplasma acidophilum*.

**Possible Involvement of Radical Intermediates?**

The mechanism of head-to-head C-C bond formation during tetraether lipid synthesis was explored by incorporation of deuterium-labeled precursors and NMR analysis by Eguchi et al. (25–27). ^1H-NMR analysis of the deuterium-labeled lipid synthesized from mevalonolactone-Δ5 in growing cells of *Haloarcus japonica* showed again that the isoprenoid was synthesized via the MVA pathway and that hydrogenation of the double bonds in the geranylgeranyl chains took place through the addition of hydrogens in a syn manner (two hydrogen molecules attacked from the same side of a double bond) (25). Similar experiments with *Methanocaldococcus jannaschii* gave similar results except that significant proton (^1H) incorporation into the C-4, C-8, C-12, and C-16 positions was observed. The protons at these positions were designated pro-S protons. It was suggested that the proton incorporation at those positions occurred regio- and stereospecifically in each phytanyl chain. This protonation pattern may well suggest the involvement of migration of the double bonds (isomerization), and this double bond migration would have to occur after the formation of the digeranylgeranylglycerol (27). Because the authors took note of the fact that this migration occurs only in archaea that have macrocyclic ether lipids with C_{40} bi-phytandiyyl chains (macrocyclic archaeol or caldarchaeol core), they considered the isomerized intermediate having a terminal methylene functionality to be potentially important for the C-C bond formation between two C_{40} isoprenoid chains. However, in contrast, deuterium-labeled archaetidic acid analogs with terminal double bonds were not incorporated into diether and tetraether lipids in *Methanothermobacter thermoautotrophicus* cells (26). The most efficient precursor was an archaetidic acid analog with a terminal isopropylidene group [(CH_3)_2CH=] for the synthesis of tetraether lipid, and this contradicts Nemoto’s result, which showed that the precursor was fully saturated archaetidylglycerol. The authors suggested the involvement of
a radical trigger reaction in the C-C bond formation in connection with the biosynthesis of a similar compound, diabolic acid (15,16-dimethyltriacontadienic acid), in a bacterium, Butyrivibrio fibrisolvens. Diabolic acid was shown to be synthesized from fully saturated [16-2H]palmitic acid or [14-2H]palmitic acid without the loss of 2H, suggesting unrelatedness with a double bond. Both experiments were done in vivo. Since in vivo experiments include inevitable ambiguity, the contradiction must be settled by appropriate in vitro experiments. However, the substrates, a detection method for the products, and the assay conditions required for in vitro experiments are all still unknown.
EVOLUTION OF MEMBRANE LIPID AND DIFFERENTIATION OF ARCHAEA AND BACTERIA

Hybrid Nature of the Phospholipid Synthesis Pathway

The enantiomeric glycerophosphate backbone, ether linkages, and isoprenoid chains are distinguishing characteristics of archaeal lipids. These structural features are synthesized in the first half of the biosynthetic pathway of the archaeal phospholipids. In bacteria, the characteristic structures of polar lipids (G-3-P backbone, ester linkages, and fatty acid chains) are also synthesized in the first half of the biosynthetic pathway. Furthermore, the orders of the reactions are arranged in a similar manner, that is, glycerophosphate backbone formation first, followed by two steps of hydrophobic chain binding.

In contrast with the core portion, several kinds of polar groups (phosphodiester-linked ethanolamine, t-serine, glycerol, myo-inositol, and choline) are membrane phospholipid constituents in all kinds of organisms. The polar groups of phospholipids are shared by organisms of all three domains. Incorporation of common polar groups into phospholipids in archaea and bacteria proceeds by the same reaction mode and is catalyzed by the homologous enzymes, that is, activation of the first lipidic intermediate by CTP and replacement of CMP by various kinds of polar compounds.

Throughout the biosynthetic pathways of phospholipids in organisms of both domains, the overall reaction sequences are analogous. Although the biosynthesis of core lipids is carried out by analogous reaction sequences, specific enzymes are at work to give the specific products of the reactions for the respective domains. On the other hand, polar head group-attaching enzymes belong to the same enzyme family and are homologous. The enzymes (at least the serine phospholipid synthases) are interchangeable, at least in terms of substrate specificities and reaction conditions. In short, the phospholipid synthesis pathway of Archaea consists of Archaea-specific reactions and Archaea-Bacteria-common reactions. This can be regarded as the hybrid nature of the phospholipid synthesis pathway.

Differentiation of Archaea and Bacteria Caused by Segregation of Enantiomeric Membrane Phospholipid, and the Evolution of Phospholipids

Koga et al. (51) considered the difference in the phospholipid backbones (G-1-P and G-3-P) to be important for the emergence of archaea and bacteria. Wächtershäuser had adopted the significance of the difference in the backbone stereostructure in the evolution of archaea and bacteria and proposed a hypothesis. According to Wächtershäuser’s hypothesis (105), precells (common ancestral cells assumed to be present before the speciation of archaea and bacteria) contain glycerophospholipids as major cell membrane constituents with racemic glycerophosphate backbone. Spontaneous physicochemical phase separation resulted in more homochiral membrane segments, and frequent and active fusion and fission of precells resulted in advanced precells with increased homochiral lipid membranes. These precells with greater homochirality and more-stable lipid membranes (with G-1-P and G-3-P backbones) would be the ancestors of archaea and bacteria, respectively. The fact that the structures of the polar head groups and the attaching enzymes are shared by archaea and bacteria suggests that the precells, before the diversification of the two domains, possessed common polar groups in their phospholipids. That is, the membrane phospholipids at that stage were glycerophospholipids with a racemic glycerophosphate backbone and several common polar head groups, as found in contemporary phospholipids. H. Morii and Y. Koga (69) speculated that the gene encoding the ancestral phosphatidylserine synthase was transferred from a gram-positive bacterium to a group of archaea based on the similarities in amino acid sequences and enzymatic properties. Now, we consider the polar head group structures and polar head group-attaching enzymes common to archaea and bacteria to be derived from a common ancestor; that is, the common phospholipids and the common enzymes are assumed to be present in precells before the differentiation of archaea and bacteria.

Differentiation of the domains would be driven by spontaneous phase separation of hetrochiral phospholipids, and the polar head groups would be carried on to the descendant archaea and bacteria. Another theory that connects the differentiation of archaea and bacteria with lipid backbone chirality has been developed by Martin and Russell (63) and by Pereto et al. (89). This theory more or less emphasizes the evolutionary significance of the enantiomeric glycerophosphate backbone of membrane phospholipid and proposes the origin of the two stereospecific glycerophosphate dehydrogenases by recruitment of the dehydrogenases already present in the universal cenancestor, which constitute the concurrent superfamilies of G-1-P dehydrogenase and G-3-P dehydrogenase.

The structures of phospholipids that constitute biological membranes have certain common features, which include having one polar head and two hydrophobic tails connected by a three-carbon backbone. This is the minimum requirement for membrane phospholipid structure. All membrane lipids possess this structure. This is presumed to be the overarching uniformity in the polar lipid structure (Fig. 12). Actual phospholipids are diverse in structure within the limited range of this structural uniformity. Hydrocarbon chains and linkages between a glycerophosphate backbone and hydrocarbon chains may be diverse; ether, ester, and alk-1’-enyl ether linkages (plasmalogen) are all possible. In a rare case, the C-C bond as a connection linkage of the two parts is possible in the long-chain 1,2-diol lipid in Thermomicrobium roseum (60). In this case, glycerol lipids are completely absent and the first three carbons with two hydroxy groups of the long-chain diol play a role in the backbone of the lipid. Hydrocarbon chains may be straight fatty acid or highly methyl-branched isoprenoids. Either stereoconfiguration of a glycerophosphate backbone is possible for formation of a membrane. The stereostructure of the glycerophosphate backbone, one example of the diversity of lipid structures, became a trigger of speciation in Archaea and Bacteria. At the time of the segregation of the chiral core lipid, both descendants had common polar groups. The fact that present-day membrane phospholipids have common polar groups is thus a matter of course. Discussions put forward by Wächtershäuser (105) and Koga et al. (51) considered only the differences in the stereostructure of glycerophosphate backbones. However, the membrane lipids in ancestral precells must have carried polar head groups as well as core lipids. They did not address the issue of the polar groups. Now, this issue of polar head groups in the membranes of precells has been addressed by studies of the structure and biosynthesis of...
The present paper has reviewed in vivo and in vitro studies conducted on polar lipid biosynthesis in archaea, along with a phylogenetic analysis of biosynthetic enzymes. Although in vivo experiments are an efficient means of garnering insight into completely unknown mechanisms, sometimes in vivo results are complicated by unexpected living phenomena (such as metabolisms). One such example is the subject of the mechanism of G-1-P structure formation. Whereas in vitro experiments can to a degree compensate for the vulnerable points of phospholipids of archaea compared with their bacterial counterparts.

**FUTURE SUBJECTS OF LIPID BIOSYNTHESIS IN ARCHAEA AND CONCLUDING REMARKS**

The present paper has reviewed in vivo and in vitro studies conducted on polar lipid biosynthesis in archaea, along with a
in vivo experiments, it is important to exercise caution that a detected reaction is confirmed to be physiologically functional in living organisms. We have learned such lessons on the strong and weak points of the two methodologies from the reconstruction of the history of biosynthetic research of archaeal polar lipids traced in this review.

Studies of the structures and biosynthesis of archaeal polar lipids and comparisons with bacteria have afforded insight into early cellular evolution, especially the early differentiation of lipids and comparisons with bacteria have afforded insight into polar lipids traced in this review.

struction of the history of biosynthetic research of archaeal lipids in living organisms. We have learned such lessons on the strong and weak points of the two methodologies from the reconstruction of the history of biosynthetic research of archaeal polar lipids traced in this review.

Many problems concerning lipid biosynthesis in archaea remain to be investigated. Although the general main pathway of phospholipid biosynthesis has been outlined in vitro, the in vitro biosynthesis of ethanolamine-, glycerol-, and myo-inositol-containing phospholipids has not been determined. Because the final products of most archaeal polar lipids are fully saturated, there should be saturation (hydrogenation) steps somewhere after the formation of unsaturated archaeal phospholipids. Studies of the mechanism for the hydrogenation reaction are under way, as the result of the discovery of a hydrogenating enzyme. This is also a unique feature of archaeal polar lipid biosynthesis that is not seen in bacteria. The mechanism of tetraether lipid biosynthesis, i.e., head-to-head condensation of the isoprenoid chain at the methyl ends, must be a novel reaction. It is, as mentioned already, one of the most challenging subjects in general biochemistry. In addition, most enzymes in the pathway have neither been purified nor characterized. Finally, while many relevant genes have been detected from database searches, these genes have not been cloned.

Hydroxyarchaeol, cyclic archaeol, cyclopentane ring-containing caldarchaeol, and H-shaped caldarchaeol may also be synthesized by novel mechanisms and hence be worthy of study. Additionally, how the polar groups of the lipids unique to archaea (e.g., aminopentanetriols, glucosaminylinositol, and glucosylinositol) are synthesized is a remaining challenge. However, these diverse problems may be too-particular sub-questions.

The purification and characterization of relevant enzymes, cloning and sequencing of the genes encoding the enzymes, phylogenetic studies of the enzymes, and comparative studies of the structure and synthesis of the lipids of archaea and bacteria all show great promise for yielding novel and valuable insights into biological lipids and membranes.

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REFERENCES

eeryl phosphate synthase—puriﬁcation and characterization of the ﬁrst path-
phate synthase from Archaeoglobus fulgidus is a class II aldolase. Biochemistry 39:12435–12439.
drogenase. Protein Eng. 15:687–695.
chemistry 21:595–599.
25. Eguchi, T., M. Morita, and K. Kamakura. 1998. Multigram synthesis of mevalonolactone-d(9) and its application to stereochemical analysis by H-1 NMR of the saturation reaction in the biosynthesis of the 2,3-di-O-
26. Eguchi, T., Y. Nishimura, and K. Kamakura. 2003. Importance of the isopropyenol terminal of geranylglycerol group for the formation of tet-


